

Putting plant resistance traits on the map: a test of the idea that plants are better defended at lower latitudes

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• It has long been believed that plant species from the tropics have higher levels of traits associated with resistance to herbivores than do species from higher latitudes. A meta-analysis recently showed that the published literature does not support this theory. However, the idea has never been tested using data gathered with consistent methods from a wide range of latitudes.

• We quantified the relationship between latitude and a broad range of chemical and physical traits across 301 species from 75 sites world-wide.

• Six putative resistance traits, including tannins, the concentration of lipids (an indicator of oils, waxes and resins), and leaf toughness were greater in highlatitude species. Six traits, including cyanide production and the presence of spines, were unrelated to latitude. Only ash content (an indicator of inorganic substances such as calcium oxalates and phytoliths) and the properties of species with delayed greening were higher in the tropics.

• Our results do not support the hypothesis that tropical plants have higher levels of resistance traits than do plants from higher latitudes. If anything, plants have higher resistance toward the poles. The greater resistance traits of high-latitude species might be explained by the greater cost of losing a given amount of leaf tissue in low-productivity environments.

Introduction

The idea that there is a latitudinal gradient in traits associated with resistance to herbivores, where plants from low latitudes are more strongly defended against herbivores than are plants from high latitudes, is very widely accepted (Schemske *et al.*, 2009). The dominant theory is that lowlatitude species experience more intense herbivory than do species at higher latitudes, and have thus been under selective pressure to evolve higher levels of resistance traits (Dobzhansky, 1950; MacArthur, 1972; Coley & Aide, 1991; Coley & Barone, 1996; Van Alstyne *et al.*, 2001). These ideas are central to our understanding of global patterns in species richness and the factors that shape latitudinal gradients in plant traits.

Some studies have provided support for the idea that plants from lower latitudes have higher levels of traits that are thought to confer resistance to herbivores. For example, studies have reported latitudinal gradients in the proportion of species that contain alkaloids (Levin, 1976; Levin & York, 1978), latex (Lewinsohn, 1991), and extrafloral nectaries (Pemberton, 1998), and others have shown latitudinal gradients in leaf toughness and tannin and total phenolic concentrations both within (Siska et al., 2002) and across species (Coley & Aide, 1991; Hallam & Read, 2006). However, many studies do not show higher levels of resistance traits at lower latitudes, either within species (Lesage et al., 2000; O'Neill et al., 2002; Gaston et al., 2004; Stark et al., 2008; Adams et al., 2009; Martz et al., 2009) or across species (Ardon et al., 2009; Graca & Cressa, 2010; Steinbauer, 2010). A recent review found that only nine of the 56 latitudinal comparisons of plant chemical traits available in the literature showed higher levels of resistance traits at lower latitudes (Moles et al., 2011). Meta-analysis showed that the average effect size from these studies was indeed significantly different from zero, but in the opposite direction from that predicted by much of the literature. On average, chemical resistance traits were c. 40% higher levels at the high-latitude end of the comparisons (Moles et al., 2011). A review of the literature also failed to support the idea that physical resistance traits would be greater in lower latitude plants. Only 13 of the 25 latitudinal comparisons of physical resistance traits found higher levels of resistance at lower latitudes, and meta-analysis showed no significant difference between the average effect size and zero (Moles et al., 2011). Of course, the result of this meta-analysis does not invalidate any of the studies that have shown latitudinal gradients in resistance traits - it simply provides an estimate of the combined weight of evidence on the question of whether there is a latitudinal gradient in traits associated with resistance to herbivores.

On the surface, it appears that we should immediately reject the idea that plants at low latitudes have higher levels of traits that confer resistance to herbivores. However, Moles *et al.* (2011) caution that the data on which their analysis is based are far from perfect, with most studies contrasting sites in just two or three regions rather than from a range of latitudes, applying imperfect or inconsistent methods, spanning only a fraction of the latitudinal gradient and/or including relatively small numbers of species. To provide a conclusive answer to the question of whether

there is a general latitudinal gradient in plant resistance traits, we need a study that applies consistent methods to a range of species from sites spanning as much of the latitudinal gradient as possible. The main aim of the present paper was to report on such a study.

We quantified a broad array of physical and chemical traits that are thought to affect vulnerability to herbivores (putative resistance traits), across 301 species from 216 genera and 85 families. These were the four most abundant species (in terms of leaf area displayed) from each of 75 relatively natural sites from all around the world, including rainforests in the Republic of Congo, Panama, China and Mexico, deserts in Israel, Australia and the USA, tundra in Greenland and Alaska, savannas in Zambia, South Africa and Australia, and temperate forests in Sweden, the USA and Australia (Fig. 1). Our aim was to determine whether plants from low latitudes have higher levels of resistance traits than do plants from high latitudes.

Materials and Methods

Site location and species selection

We worked at 75 study sites, distributed from 74.5°N to 51.5°S (Fig. 1; Supporting Information Table S1). Sites were selected to sample the dominant vegetation types at a wide range of latitudes. Details of site selection are given in Methods S1, and a list of sites is presented in Table S1, but the primary criterion was that the levels of herbivory, disturbance regime and plant community composition should be relatively natural (i.e. as close as possible to those with which the plant traits we are measuring are thought to have evolved). At each site, we sampled the four most abundant species (exceptions are outlined in Methods S1).

Leaf sampling

We sampled fully expanded photosynthetic units for each species (usually leaves or leaflets, but occasionally photosynthetic stems or phyllodes). All of these photosynthetic units are henceforth referred to as 'leaves', for convenience. Leaves from at least five mature, outwardly healthy individuals of each study species were sampled as close to the peak growing season as possible. Full details of leaf selection are given in Methods S1.

A total of at least 40 g of fresh leaves was collected from at least five individuals of each species, and placed in paper bags ready for oven-drying, and an additional three fresh leaves from each of five plants of each species were placed in plastic bags with damp tissue, and stored in a cooler or refrigerator until they could be measured.

Leaf traits

We did not investigate the precise nature of the chemistry for each species, or try to account for the feeding preferences of herbivores in different parts of the world. Instead, we tried to select a wide range of broad physical and chemical traits that would capture as much information about the species' likely vulnerability to herbivores as possible. These traits are referred to as 'resistance traits' for convenience throughout the paper. These traits are among the most commonly identified traits involved in resistance, and it is generally believed that these traits increase plant fitness in the presence of herbivores (but see Carmona et al., 2011). Most of these traits have multiple functions, and, while our correlative approach shows global patterns in traits, it cannot resolve the causes of latitudinal gradients. However, we see obtaining a firm quantification of the global patterns as an important first step in understanding global patterns



Fig. 1 World map, showing study site locations. Some points represent more than one site (this occurs where two or more different ecosystems were close in geographic space).

in plant–animal interactions. A rationale for the inclusion of each trait is given in the following sections, along with an explanation of our measurement techniques.

Leaf toughness and size We began by measuring physical toughness, an extremely important form of resistance against herbivores (Choong et al., 1992; Turner, 1994; Hanley et al., 2007; Clissold et al., 2009). We used the leaf slicing machine designed by Wright & Cannon (2001) to measure the force required to push a blade through 10 fresh leaves (mean force of fracture (N); a detailed description is given in Wright and Cannon, 2001). Our data set for force of fracture is smaller than for other traits, because of the difficulty of transporting the instrument to the different sites to use on fresh leaf material. We therefore complemented the fracture data with measures of specific leaf area (SLA), which is negatively related to force of fracture (standardized major axis slope = -0.76; $R^2 = 0.53$). We used a flat-bed scanner to scan 15 fresh leaves, with a ruler or grid for scale. Images were analyzed using IMAGE-J (National Institute of Health, Bethseda, MD, USA) to calculate leaf area. These leaves were dried at 55-65°C (the vast majority at 55°C), for 24-48 h, and weighed for dry mass. Specific leaf area (SLA) was calculated by dividing mean leaf area by mean dry mass.

Although leaf size is not a resistance trait *per se*, it does affect the vulnerability of the leaf to herbivores. Larger leaves have been shown to experience greater degrees of herbivory within a habitat (Moles & Westoby, 2000), probably because of their greater apparency to herbivores (Feeny, 1976), and/or because of their greater expansion times (Moles & Westoby, 2000).

For the remaining traits, we used dried, ground leaves. Samples were oven-dried at temperatures between 55 and 65°C (the vast majority at 55°C). With very few exceptions, samples were placed in a drying oven within 6 h of collection. No method of drying leaf tissue is optimal and every method (including freeze-drying and prolonged air-drying) may change plant chemistry. We chose oven-drying at low temperatures as a reasonable compromise between speed of dehydration (and so reduction in plant enzyme activity) and ability to be performed consistently over all sites. A key measure in our study was polyethylene glycol (PEG)-binding capacity, a measure of tannin activity, and this measure has been found not to differ between oven-dried and freezedried material (Silanikove *et al.* 1996).

Dry samples were packed in plastic sample vials, and transported to Sydney, Australia. Samples from outside Australia were gamma-irradiated at 50 kGray, according to quarantine requirements (this does not affect the gross chemical composition of the samples, except by rendering them biologically sterile). All samples were ground to pass a 1-mm sieve on an Udy Cyclone Sample Mill (Udy Corporation, Fort Collins, CO, USA), and stored refrigerated in the dark until analyses could be performed. All chemical analyses except that of ash content were performed in duplicate. Analyses of any pair of samples whose values were too different (usually a CV > 5%) were repeated in duplicate.

Cyanogenic glycosides The ability to release hydrogen cyanide in response to cell damage is a widespread resistance trait that is known to be effective against a wide range of invertebrate and vertebrate herbivores (Ballhorn et al., 2010). To test for cyanogenesis, we added 150 µl of phosphate buffer (pH 5.0; 0.1 M) to 0.20 g (± 0.01 g) of dried, ground leaf in a sealed Vacutainer (Becton-Dickinson, Franklin Lakes, NJ, USA) with a strip of Feigl-Anger paper (which turns blue in the presence of cyanogenic glycosides; Feigl & Anger, 1966) suspended from the top. If no activity was observed within 24 h, the assays were stopped (because of the possibility of false results from cyanogenic bacteria). New assays were initiated for all species that showed a negative response, in which 1.12 units $ml^{-1} \beta$ -glucosidase from almond (Prunus dulcis) (Sigma-Aldrich) was added to the buffer to determine whether each species could release cyanide if enzymes were supplied from a source other than the plant (for example, if a glucosidase was available during digestion inside a herbivore). This method is the same as that used by Marsh et al. (2007). Eleven species responded positively in the presence of the glucosidase, while nine species responded positively with buffer only. We report the results from the trials with glucosidase throughout.

We used near-infrared reflectance spectroscopy (NIRS) to predict some values for the remaining chemical traits (Foley *et al.*, 1998). We followed the standard procedures recommended by the American Society for Testing and Materials for all quantitative NIRS analyses (A.S.T.M., 1995; Foley *et al.*, 1998). In brief, after collecting duplicate spectra on a FOSS NIR-Systems 6500 spectrophotometer (FOSS, Hillerød, Denmark), we selected 100 species based on their spectral variability. We conducted full chemical analyses on these samples and then developed partial least squares regression models to relate chemical and spectral variation. After validation, we used these models to predict the chemical composition of unmeasured samples from their spectra. Details of these models can be found in Methods S1 and Table S2.

Lipids We measured total lipid content, to quantify resistance traits such as oils (including terpenes), cuticular waxes, and resins. Several studies have shown that these traits deter or otherwise negatively affect herbivores, probably through a combination of physical and chemical effects (Lincoln, 1985; Peeters, 2002; Jones *et al.*, 2003; Marko *et al.*, 2008). For instance, cuticle thickness was negatively correlated with densities of a range of invertebrate herbivores, including sessile phloem feeders, rostrum chewers, and external chewers (Peeters, 2002). Juniper (*Juniperus Communis*) individuals with higher concentrations of essential oils are less damaged by herbivores (Marko *et al.*, 2008), and kangaroos avoid plant species with volatile essential oils (Jones *et al.*, 2003). Finally, the growth of butterfly larvae was inhibited by high concentrations of leaf resin in *Diplacus aurantiaeus* (Lincoln, 1985).

To measure lipid content, we added 10 ml of petroleum spirit (bp 40–60°C; also known as petroleum ether) to 1 g (\pm 0.01 g) of dried, ground leaf sample, in a pre-weighed 10-ml centrifuge tube. Samples were shaken vigorously, placed in a sonicator for 20 min and then left to stand for 2 h. Previous trials showed that longer periods of mixing and standing did not increase weight loss from the sample. Samples were centrifuged at 4000 g for 5 min and then the lipid extract was decanted quantitatively. The remaining sample was oven-dried at 50–60°C for 5–6 d and reweighed to calculate the lipid loss. Ether-extractable lipids have previously been shown to be negatively correlated with herbivore preferences across a range of herbivores (Bryant & Kuropat, 1980).

Tannins Tannins have been found to deter feeding in a range of herbivores, including mammals, slugs and insects (Furstenburg & Vanhoven, 1994; Fritz *et al.*, 2001; Roslin & Salminen, 2008). In mammals, tannins reduce protein and dry matter digestibility, cause endogenous nitrogen loss, and can cause damage to the gastrointestinal tract, kidney and liver (Shimada, 2006; Spalinger *et al.*, 2010). Tannins have also been shown to reduce metabolic and growth efficiency in invertebrates (Roslin & Salminen, 2008). There is also increasing evidence that the oxidative capacity of tannins might be an important component of their role as resistance traits (Salminen & Karonen, 2011).

We employed PEG-binding capacity, determined using Silanikove et al.'s (1996) PEG-binding assay, as a measure of tannins. We chose the PEG-binding assay over more traditional colorimetric assays of phenolics for two reasons. First, it measures the degree to which tannins bind the plant protein. That is, it measures a functional trait that is relevant to animals (particularly mammals), rather than quantifying the concentration of a suite of chemicals that can have varying effects on herbivores. Second, the PEGbinding assay does not rely on extraction of tannins from the plant matrix and their subsequent quantification using external standards. These two issues are major limitations of most tannin analyses and preclude comparison of data for widely differing taxa because different tannins produce different chromophores at similar concentrations, making interpretation of colorimetric methods problematic across different species without detailed knowledge of their chemistry (Mueller-Harvey, 2006).

PEG-binding capacity was measured by mixing 0.50 g of dried ground leaf with 7.5 ml of a solution containing

33.33 g l⁻¹ PEG 4000 spiked with [¹⁴C]-labeled PEG 4000. Samples were incubated for 24 h and centrifuged, and 75 μ l of supernatant was combined with 10 ml of scintillant and counted (as in DeGabriel *et al.* 2008).

Nitrogen digestibility, carbon (C) : nitrogen (N) ratio and N fixation Leaves with low N content and/or high C : N ratios are generally considered lower quality food for herbivores (Mattson, 1980; Schmitz, 2008). Although N content is not a resistance trait, we consider it here because herbivores have been shown to balance N intake against the defensive properties of their food (Bryant & Kuropat, 1980; Galimuhtasib *et al.*, 1992). Leaf C and N contents were measured using a LECO C : N : S analyser (LECO, St. Joseph, MI, USA) at the Environmental Analysis Laboratory at Southern Cross University, Lismore, Australia.

We determined *in vitro* N digestibility using the method of DeGabriel *et al.* (2008). The assay involved digesting the samples in porous bags (Ankom F57; Ankom Technology, Macedon, NY, USA), first with pepsin (24 h) and then with cellulase (48 h). We analyzed the residue at the end of the assay to quantify the digestibility of the N (%). We were interested in measuring the effects of tannins on available N, so at the start of the digestion we incubated two bags per sample for 24 h either with 33.3 g l⁻¹ PEG 4000 (in 0.05 M Tris-BASE buffer) or with buffer alone (both 25 ml per sample). We then thoroughly washed the bags before drying them to constant mass at 50°C and weighing them.

We scored each plant for presence/absence of the ability to fix N, based on information from the published literature.

Ash We measured ash content, in order to gain information about resistance traits such as silica-based phytoliths and calcium oxalates. Calcium oxalate is present in most plant families, and is the most abundant insoluble mineral in plant tissue, accounting for 3-80% of plant dry mass (Franceschi & Nakata, 2005; Korth et al., 2006). Silica is also widespread and abundant in plant tissues, especially in grasses, which are typically 2-5% silica (Massey et al., 2006). Both silica and calcium oxalates reduce feeding by a range of herbivores, including insects and mammals (Djamin & Pathak, 1967; Galimuhtasib et al., 1992; Ward et al., 1997; Korth et al., 2006; Massey et al., 2006; Hanley et al., 2007). These minerals increase the abrasiveness of leaf material, and reduce herbivore growth rates and digestion efficiency (Korth et al., 2006; Massey et al., 2006). There was a highly significant positive correlation (P = 0.001; $R^2 = 0.50$) between log₁₀ ash content and log₁₀ silica content across 27 terrestrial species (data analyzed were from Lanning & Eleuterius, 1985).

To calculate ash content, 1.00 g of dry sample was combusted at 600°C for 12 h before weighing the residual ash.

Other traits We used observation in the field, published records and/or local knowledge to score each species for possession of a range of additional traits: delayed greening, latex, hair on adult and/or juvenile leaves, extrafloral nectaries, and spines. Each of these traits is thought to reduce damage by herbivores. Delayed greening (possession of red, white or pink young leaves that have low chlorophyll content) is associated with a low N content, which may reduce the attractiveness of young foliage to herbivores and reduce the cost of herbivory on juvenile leaves (Kursar & Coley, 1992). Latex is present in over 20 000 plant species, and several lines of evidence suggest that it acts to deter chewing herbivores, particularly invertebrates (Agrawal & Konno, 2009). The presence of hair on leaves is thought to make it more difficult for invertebrates to access leaf tissue, and has been shown to decrease losses to herbivores (Moles & Westoby, 2000; Hanley et al., 2007). Extrafloral nectaries are found in over 90 plant families, and the omnivorous invertebrates they attract (typically ants) attack herbivorous insects, thus increasing the host plant's survival and/or reproductive success (Ness et al., 2009). Spines (including thorns) have been shown to be an effective form of resistance against a range of herbivores, particularly mammals (Hanley et al., 2007).

Statistics

Variables were log10-transformed as appropriate. We used linear mixed-effects models (Venables & Ripley, 2002) for each of the traits described in the preceding sections, with a fixed-effect term for latitude and a random effect term for site. We included an interaction term for hemisphere in the analyses, to allow for different relationships in the Northern and Southern Hemispheres. The random-effects term for site was included to account for site-to-site variation in leaf traits that was not explained by latitude. This term also allowed us to determine what percentage of the unexplained variation lay within vs across sites. Models were fitted using restricted maximum likelihood via the R package lme4 (Bates *et al.*, 2008). With data for multiple sites, R^2 for a term in the model (fixed or random) was calculated by the usual method (sequential reduction in residual sum of squares on addition of the term), but adding fixed-effects terms to the model before the random-effects term. Binary variables (such as presence or absence of spines) were analyzed using logistic regression with a random site term via lme4 (Bates et al., 2008).

We also quantified the relationships between species' cover (absolute and relative) at each site and each of the resistance variables, to determine the extent to which our selection of the four most abundant species might have affected our results. Absolute cover values are estimates of the leaf area index for each species in a series of three to seven quadrats, while relative cover divides the cover of the focal species by the total cover in the quadrats. Both cover metrics were log₁₀-transformed before analysis for normality, and trait variables were log-transformed where appropriate, as for previous analyses. Continuous traits were analyzed with ordinary regressions, while binary traits were analyzed with logistic regressions, both in PASW 18 (formerly SPSS, IBM Corporation, Somers, NY, USA).

Results

The PEG-binding capacity (a measure of tannins) of species in the tropics was lower than that of species at high latitudes $(P = 0.047; R^2 = 0.02;$ Fig. 2a). Species found > 60° from the equator had a mean PEG-binding capacity 1.6 times higher than that of species growing in the tropics (7.5 vs 4.7 g 100 g⁻¹, respectively). The highest PEG-binding capacity was found in *Eucalyptus miniata*, from Darwin, Australia (30 g 100 g⁻¹), followed by *Crassula rupestris* from near Stellenbosch, South Africa (22 g 100 g⁻¹), and *Pistacio lentiscus* from Israel (21 g 100 g⁻¹).

Our analyses of *in vitro* N digestibility support the idea that tannins are decreasing the availability of N to herbivores at high-latitude sites. There was no latitudinal gradient in N digestibility in the absence of PEG (P = 0.17), but when PEG (which binds to tannins, rendering them inactive) was added to the samples before digestion, high-latitude species had significantly higher N digestibility (P = 0.04; $R^2 = 0.02$; Fig. 2b).

There was a striking latitudinal gradient in lipid concentration ($R^2 = 0.21$; Fig. 2c). Species found > 60° from the equator had a mean lipid concentration more than twice as high as that of species growing in the tropics (5% vs 2.1%). The highest lipid concentration in our samples was 9.9% in *Picea mariana* from Alaska, followed by three species of *Eucalyptus* from near Adelaide in Australia that ranged from 9.8 to 8.7% lipid. One possible explanation for the higher lipid concentrations at high latitudes is that the lipids might protect cells from freezing damage. Plants from colder climates tend to have higher unsaturation of lipids (Badea & Basu, 2009), but there has been little focus on the relationship between total lipid concentration and cold tolerance.

Ash content was significantly higher in tropical species $(P = 0.014; R^2 = 0.03;$ Fig. 2d). We wondered whether the higher ash content in tropical species might result from the inclusion of several grass species from tropical savannas, as species in the *Poales* are known to have high silicon content (Hodson *et al.*, 2005). We therefore added a term for grass/nongrass to the analysis. There was no significant interaction between this term and latitude (P = 0.42), indicating that the slope of the relationship was not significantly different between grasses and nongrasses. However, as expected, grasses did have a significantly higher intercept than did nongrasses (P = 0.003).



Fig. 2 Relationships between latitude and resistance traits. (a) Polyethylene glycol-binding capacity (a measure of tannis), (b) N digestibility in the presence of polyethylene glycol, which renders tannis inactive, (c) Lipid concentration, (d) Ash content, (e) Force of fracture, (f) Specific leaf area, (g) C : N ratio, (h) Leaf area, (i) Delayed greening, (j) Possession of spines, (k) Presence of hair on juvenile leaves. Each point represents one species at a site. Symbols represent growth forms: trees, squares; shrubs, triangles; herbs (including grasses), circles; climbers, diamonds. In (d), grasses are shown as open circles while all other taxa are closed. Relationships for continuous traits are from linear mixed-effects models, and are plotted with Northern and Southern Hemispheres separately (Northern Hemisphere, positive latitudes; Southern Hemisphere, negative latitudes). Relationships for binary traits are from random-effects logistic regressions. The line in (i) shows the probability of a species from a given latitude possessing delayed greening (scale on right *y*-axis). As there was no significant difference in the slope of the relationship between the two hemispheres for any binary trait, these relationships are plotted with absolute latitude (distance from equator) on the *x*-axis. ODM, over dry matter.

Leaf toughness, measured as force of fracture, decreased toward the pole in the Northern Hemisphere, but increased toward the pole in the Southern Hemisphere ($R^2 = 0.36$; Fig. 2e). There was no relationship between SLA and latitude in the Northern Hemisphere (P = 0.74), but there was a significant relationship in the Southern Hemisphere, with SLA increasing toward the equator (P < 0.001; $R^2 = 0.10$; Fig. 2f; the SLA at 60°S (4.44 mm² mg⁻¹) was only 29% that at the equator (15.3 mm² mg⁻¹)). Taking force of frac-

ture and SLA together, tropical leaves seem to have slightly less physical resistance to herbivores than do leaves from high latitudes. A likely explanation for the difference in SLA and force of fracture between the Northern and Southern Hemispheres is the greater prevalence of deciduous leaves in the Northern Hemisphere, and thus longer leaf lifespans in the Southern Hemisphere.

There was a substantial latitudinal gradient in C : N ratio $(R^2 = 0.12; \text{ Fig. 2g})$, with species growing > 60° from the

equator having a mean C : N ratio 1.7 times higher than that of species growing in the tropics. This gradient was generated by both lower concentrations of C ($R^2 = 0.07$) and higher concentrations of total N ($R^2 = 0.12$) in tropical leaves. We also found that significantly more of the dominant species in tropical communities are able to fix N (P = 0.04; $R^2 = 0.04$). These data, combined with those for tannins and N availability, suggest that tropical species have higher nutritional value, and are thus likely to be more attractive to herbivores.

There was a strong relationship between latitude and leaf area ($R^2 = 0.27$; Fig. 2h), with leaves at the equator averaging 43 times the size of leaves at 60°N. Although the existence of a latitudinal gradient in leaf size has been known for some time (e.g. Webb, 1959), our study is the first global-scale quantification of this important trait.

Delayed greening was present in a significantly higher proportion of species toward the tropics (P = 0.016; $R^2 = 0.05$; Fig. 2i). The fact that a high proportion of tropical species have delayed greening has been noted before (Coley & Kursar, 1996; Dominy *et al.*, 2002), but, contrary to some previous reports (Coley & Barone, 1996), delayed greening was not restricted to tropical species. Delayed greening seems mostly to be a mechanism for reducing the amount of N lost to herbivores, rather than a resistance trait *per se*, but this remains one trait whereby tropical species are less vulnerable to herbivores than are their temperate counterparts.

There was no significant relationship between latitude and the presence of spines (Fig. 2j), hairs on either juvenile or adult leaves (Fig. 2k), extrafloral nectaries, or latex (all P > 0.2; Table S3). Eleven of the 286 species tested positive for cyanogenic capacity, but this ability was not significantly related to latitude (Fig. 2l). Nonsignificant results for some traits (especially extrafloral nectaries and latex) might be a consequence of the small number of species in our data set that possessed the trait (Table S3).

Data considerations

To determine whether any of the relationships between latitude and resistance traits investigated here were affected by our inclusion of species with different growth forms, we conducted a series of analyses including terms for growth form (tree, shrub, herb (including grasses) or climber). In no case was there a significant interaction between growth form and latitude (all P > 0.1), indicating that the slopes of the relationships between resistance traits and latitude were not significantly different among species with different growth forms. In only two cases was the main effect of a growth form term significant (P < 0.05). Thus, in most relationships, the intercepts of the lines for species with different growth forms did not differ significantly. The exceptions were that herbs had a significantly lower intercept in the relationship between latitude and C concentration (P = 0.003), while shrubs had a significantly lower intercept in the relationship between latitude and log_{10} leaf area (i.e. they had smaller leaves at a given latitude; P = 0.02). Despite these minor differences, it is clear that the relationships between resistance traits and latitude are neither obscured nor artificially strengthened by the inclusion of species with different growth forms.

We quantified the relationship between the resistance traits and the relative and absolute cover of each species, to determine whether our selection of the four most abundant species at each site might have influenced our results (because these abundant species might make up a higher proportion of the total cover at high-latitude sites than at low-latitude sites; Kleidon et al., 2009). Only four out of 34 relationships between resistance traits and cover were significant at P < 0.05, and the highest R^2 was 0.031 (Table S4). Thus, the relationships between abundance and plant resistance traits are too weak to be driving our findings regarding the latitudinal gradient in resistance. The significant relationships were a positive relationship between absolute cover and leaf area (P = 0.008), negative relationships between relative cover and both lipid content (P = 0.048) and specific leaf area (P = 0.026), and a positive relationship between relative cover and C: N ratio (P = 0.021).

We performed phylogenetic analyses on all of the relationships investigated in this study (Methods S1, Table S5). The phylogenetic and cross-species analyses gave qualitatively similar results for 14 of 17 traits. The three exceptions were relationships that were significant in crossspecies analysis but nonsignificant in phylogenetic analysis. Crucially, there was no trait for which cross-species analyses showed no significant relationship between latitude and resistance or significantly higher resistance traits toward the poles where the phylogenetic analysis showed higher resistance traits at lower latitudes. That is, the phylogenetic analyses rule out the possibility that the expected relationship between resistance traits and latitude (higher levels of resistance traits in the tropics) is being obscured by phylogenetic history.

One of the strengths of our global sampling is that we have data from six different continents. However, different trends in the different continents could have obscured global relationships between latitude and resistance traits. We therefore ran models that included a term for continent. Some of the terms for continents were significant (Table S6). However, the overall significance and direction of the relationships between latitude and resistance traits were generally similar between models that included a term for continent and those that analysed data at the cross-continental scale. The exceptions were N fixation and PEG-binding capacity, which moved from being marginally significant (P = 0.04 and P = 0.047, respectively) to being marginally nonsignificant (P = 0.08 and P = 0.075, respectively), and leaf area, which became nonsignificant (P = 0.19) once a term for continent had been included (Table S6).

The relationships between latitude and plant resistance traits had relatively low R^2 values (mean R^2 for continuous traits = 0.16; max = 0.36; min = 0.02). However, a great deal of the unexplained variation lay at the within-site level (mean R^2 unexplained within sites = 0.45; max = 0.64; min = 0.31). That is, much of the unexplained variation was between coexisting species, and thus could not possibly be explained by latitude. Latitude explained an average of 29% of the between-site variation in plant resistance traits.

Discussion

Our data do not support the idea that plants from low latitudes have higher resistance to herbivores than do highlatitude species. If anything, the trend appears to be for greater resistance traits at higher latitudes. We measured six traits for which high-latitude species have higher levels of resistance than do low-latitude species: PEG-binding capacity, lipid concentration, leaf size, specific leaf area, force of fracture, and C : N ratio. We measured six resistance traits for which there is no significant relationship between resistance traits and latitude: ability to synthesize cyanide, presence of hairs on juvenile and mature leaves, extrafloral nectaries, spines, and latex. We measured only two resistance traits for which low-latitude species have higher levels of resistance than do high-latitude species: delayed greening and ash content. Our findings are based on the most comprehensive and consistent data set collected to date.

It is possible that, if we had measured additional traits, we might have found more evidence of higher resistance to herbivores at lower latitudes. However, it would take a lot of additional traits behaving differently from those we did measure to change the overall conclusion of our study, and we have measured a selection of the most important known resistance traits.

The results from the present study, which used consistent methods to quantify latitudinal gradients in 14 resistance traits in 301 species from 75 sites world-wide, are consistent with the findings of the recent meta-analysis of data from the published literature (Moles *et al.*, 2011). The congruence of evidence from a large-scale empirical study and a comprehensive synthesis of data from the literature strongly suggests that traditional ideas in this field need to be overturned.

The main reason ecologists predicted stronger resistance traits at lower latitudes is that low-latitude species were thought to experience more intense herbivory than species at higher latitudes, thus being under selective pressure to evolve higher degrees of resistance (Dobzhansky, 1950; MacArthur, 1972; Coley & Aide, 1991; Coley & Barone, 1996; Van Alstyne *et al.*, 2001). However, it is not clear

that herbivory is actually more intense at lower latitudes. Meta-analysis of data from the literature did not support the idea that there is a latitudinal gradient in herbivory (Moles et al., 2011). This result is consistent with findings from palaeoecology. Damage diversity on fossil leaves tracks mean annual temperature tightly through time, suggesting a greater diversity of herbivores in warmer times (Currano et al., 2010). However, damage frequency is much more weakly related to mean annual temperature, and this correlation becomes nonsignificant once serial autocorrelation is removed (Currano et al., 2010). If there is no latitudinal gradient in herbivory, then the fact that plant resistance traits are not stronger at lower latitudes is considerably less surprising. Performing a field study that quantifies the latitudinal gradient in herbivory using appropriate and consistent methods that account for differences in leaf lifespan at sites at a wide range of latitudes around the world is a top priority for understanding patterns in plant-animal interactions through both space and time.

Both Moles *et al.*'s (2011) meta-analysis and the present empirical study show that, if anything, plant resistance traits actually tend to be higher at high latitudes. One possible explanation is that the cost of losing leaves in the relatively high-productivity environments at low latitudes might be lower than the cost of losing leaf area at higher latitudes, where productivity is lower. A latitudinal gradient in the cost of losing leaf area would favor higher levels of resistance in low-productivity/short growing-season environments, including high-latitude systems. This idea is a basic extension of the resource availability hypothesis (Coley *et al.*, 1985; Hallam & Read, 2006) and Janzen's suggestion that plants should produce better protected leaves in lowproductivity environments (Janzen, 1974).

Another possibility is that the latitudinal gradients in leaf chemical and physical traits are driven not by herbivory, but by abiotic conditions. Many traits are known to have dual roles in protection against herbivores and the environment. For instance, scleromorphy protects leaves from damage from the abiotic environment, as well as from herbivores (Turner, 1994). Environmental factors such as soil fertility and exposure to UV are known to influence traits such as leaf toughness, resins and the phenolic content of leaves (Wainhouse et al., 1998; Close et al., 2003; Jordan et al., 2005). If latitudinal gradients in resistance traits are driven by abiotic conditions rather than herbivory, then either abiotic factors are more important drivers of selection on leaf traits than is herbivory, or degrees of herbivory do not vary as substantially across the globe as do abiotic factors. These are interesting possibilities that definitely merit attention. However, latitudinal gradients in traits such as phenolics have long been accepted as evidence for a latitudinal gradient in resistance to herbivores (e.g. Schemske et al., 2009). Moving the goalposts because the present evidence does not support traditional ideas would be disingenuous.

Our findings raise a host of interesting questions about large-scale patterns in plant resistance traits and plant– animal interactions. In the near future, we plan to quantify correlations among resistance traits, determine which environmental factors underlie latitudinal gradients in each trait, and investigate phylogenetic patterns in the evolution of plant resistance traits (including asking whether gymnosperms have higher levels of resistance traits than do angiosperms). There are also important questions that cannot be addressed with the present data, the most pressing of which is quantifying the effects of leaf longevity on plant resistance traits. We hope that the many questions raised by our findings will stimulate a burst of research on global patterns in plant–animal interactions and plant resistance traits.

Our results have implications beyond the understanding of plant–animal interactions. The world-wide distribution of compounds associated with resistance to herbivores has important implications for the global C cycle. Globally, the effect of leaf chemistry is the predominant control of decomposition rates (Cornwell *et al.*, 2008), and as such the patterns of leaf resistance traits described here may represent crucial information for understanding rates of terrestrial C cycling across latitude. One theory is that C that is not well defended and is thus consumed by herbivores is rapidly respired back to the atmosphere, while better defended tissue will have a longer residence time on the plant and as litter.

This is the first study to examine large-scale patterns in the relationships between plant cover and resistance traits across a range of species and sites. Across the 34 relationships between cover and resistance traits in this study, the highest R^2 was 0.031 and only four of these regressions were significant at P < 0.05. That is, we found no biologically relevant relationship between percentage cover and investment in resistance traits. One might have expected that resistance would increase with plant apparency and thus percentage cover (Feeny, 1976). However, there are reasons to predict null or even negative relationships. For example, there is often a trade-off between somatic growth and investment in resistance traits (e.g. Coley et al., 1985; Yamamura & Tsuji, 1995; VanDam et al., 1996; see Hanley et al., 2007 for caveats), suggesting that across species percentage cover could decline with increases in resistance traits. Alternatively, as productive sites are expected to benefit species with little investment in resistance traits, while species with greater herbivore resistance traits should dominate in unproductive sites (Herms & Mattson, 1992; Yamamura & Tsuji, 1995; see Hanley et al., 2007 for caveats), an interaction between a resource supply effect and growth-defence trade-offs (e.g. Fine et al., 2004) might lead to no relationship between percentage cover and resistance traits when sites that differ in productivity are pooled.

Our results, combined with findings from a recent metaanalysis (Moles *et al.*, 2011), strongly suggest that plants at low latitudes do not have higher levels of traits associated with resistance to herbivores than do plants at high latitudes. These findings cast serious doubt on traditional ideas about global patterns in plant resistance traits, and leave ecologists in a position to formulate new theories about the factors that shape plant strategy, species coexistence, and plant–animal interactions in different environments. Our results also suggest that we should ask whether other 'wellknown' relationships might not be supported by empirical data. There are exciting times ahead.

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References

- Adams JM, Rehill B, Zhang YJ, Gower J. 2009. A test of the latitudinal defense hypothesis: herbivory, tannins and total phenolics in four North American tree species. *Ecological Research* 24: 697–704.
- Agrawal AA, Konno K. 2009. Latex: a model for understanding mechanisms, ecology, and evolution of plant defense against herbivory. *Annual Review of Ecology Evolution and Systematics* 40: 311–331.
- Ardon M, Pringle CM, Eggert SL. 2009. Does leaf chemistry differentially affect breakdown in tropical vs temperate streams? Importance of standardized analytical techniques to measure leaf chemistry. *Journal of the North American Benthological Society* 28: 440–453.
- A.S.T.M. 1995. Standard practices for infrared, multivariate, quantitative analysis. Designation: E1655-94. In. American Society for Testing and Materials, West Conshohocken, PA, USA.
- Badea C, Basu SK. 2009. The effect of low temperature on metabolism of membrane lipids in plants and associated gene expression. *Plant Omics Journal* 2: 78–84.
- Ballhorn DJ, Kautz S, Lieberei R. 2010. Comparing responses of generalist and specialist herbivores to various cyanogenic plant features. *Entomologia Experimentalis Et Applicata* 134: 245–259.

Bates D, Maechler M, Dai B. 2008. lme4: linear mixed-effects models using S4 classes. R package version 0.999375-18 [WWW document]. URL http://lme4.r-forge.r-project.org/ [accessed on 5 August 2010].

Bryant JP, Kuropat PJ. 1980. Selection of winter forage by sub-arctic browsing vertebrates: the role of plant chemistry. *Annual Review of Ecology and Systematics* 11: 261–285.

Carmona D, Lajeunesse MJ, Johnson MTJ. 2011. Plant traits that predict resistance to herbivores. *Functional Ecology* 25: 358–367.

Choong MF, Lucas PW, Ong JSY, Pereira B, Tan HTW, Turner IM. 1992. Leaf fracture toughness and sclerophylly: their correlations and ecological implications. *New Phytologist* 121: 597–610.

Clissold FJ, Sanson GD, Read J, Simpson SJ. 2009. Gross vs. net income: how plant toughness affects performance of an insect herbivore. *Ecology* 90: 3393–3405.

Close D, McArthur C, Paterson S, Fitzgerald H, Walsh A, Kincade T. 2003. Photoinhibition: a link between effects of the environment on eucalypt leaf chemistry and herbivory. *Ecology* 84: 2952–2966.

Coley PD, Aide TM. 1991. Comparison of herbivory and plant defenses in temperate and tropical broad-leaved forests. In: Price PW, Lewinsohn TM, Fernandes GW, Benson WW, eds. *Plant–animal interactions: evolutionary ecology in tropical and temperate regions*. New York, NY, USA: Wiley, 25–49.

Coley PD, Barone JA. 1996. Herbivory and plant defenses in tropical forests. *Annual Review of Ecology and Systematics* 27: 305–335.

Coley PD, Bryant JP, Chapin FS. 1985. Resource availability and plant antiherbivore defense. *Science* 230: 895–899.

Coley PD, Kursar TA. 1996. Anti-herbivore defenses of young tropical leaves: physiological constraints and ecological trade-offs. In: Smith AP, Mulkey SS, Chazdon RL, eds. *Tropical forest plant ecophysiology*. New York, NY, USA: Chapman & Hall, 305–336.

Cornwell WK, Cornelissen JHC, Amatangelo K, Dorrepaal E, Eviner VT, Godoy O, Hobbie SE, Hoorens B, Kurokawa H, Perez-Harguindeguy N et al. 2008. Plant species traits are the predominant control on litter decomposition rates within biomes worldwide. Ecology Letters 11: 1065–1071.

Currano ED, Labandeira CC, Wilf P. 2010. Fossil insect folivory tracks paleotemperature for six million years. *Ecological Monographs* 80: 547–567.

DeGabriel JL, Wallis IR, Moore BD, Foley WJ. 2008. A simple integrative assay to quantify nutritional quality of browses for herbivores. *Oecologia*, 154: 107–116.

Djamin A, Pathak MD. 1967. Role of silica in resistence to Asiatic rice borer *Chilo suppressalis* (Walker) in rice varieties. *Journal of Economic Entomology* 60: 347.

Dobzhansky T. 1950. Evolution in the tropics. *American Scientist* 38: 209–221.

Dominy NJ, Lucas PW, Ramsden LW, Riba-Hernandez P, Stoner KE, Turner IM. 2002. Why are young leaves red? *Oikos* 98: 163–176.

Feeny P. 1976. Plant apparency and chemical defense. *Recent Advances in Phytochemistry* 10: 1–40.

Feigl F, Anger V. 1966. Replacement of benzidine by copper ethylacetoacetate and tetra base as spot-test reagent for hydrogen cyanide and cyanogen. *Analyst*, 91: 282–284.

Fine PVA, Mesones I, Coley PD. 2004. Herbivores promote habitat specialization by trees in Amazonian forests. *Science* 305: 663–665.

Foley WJ, McIlwee A, Lawler I, Aragones L, Woolnough AP, Berding N. 1998. Ecological applications of near infrared reflectance spectroscopy a tool for rapid, cost-effective prediction of the composition of plant and animal tissues and aspects of animal performance. *Oecologia* 116: 293–305.

Franceschi VR, Nakata PA. 2005. Calcium oxalate in plants: formation and function. *Annual Review of Plant Biology* **56**: 41–71.

Fritz RS, Hochwender CG, Lewkiewicz DA, Bothwell S, Orians CM. 2001. Seedling herbivory by slugs in a willow hybrid system:

developmental changes in damage, chemical defense, and plant performance. *Oecologia* **129**: 87–97.

- Furstenburg D, Vanhoven W. 1994. Condensed tannin as anti-defoliate agent against browsing by giraffe (*Giraffa camelopardalis*) in the Kruger National Park. *Comparative Biochemistry and Physiology a-Physiology* 107: 425–431.
- Galimuhtasib HU, Smith CC, Higgins JJ. 1992. The effect of silica in grasses on the feeding behavior of the prairie vole, *Microtus ochrogaster*. *Ecology* 73: 1724–1729.

Gaston KJ, Genney DR, Thurlow M, Hartley SE. 2004. The geographical range structure of the holly leaf-miner. IV. Effects of variation in host-plant quality. *Journal of Animal Ecology* 73: 911–924.

Graca MAS, Cressa C. 2010. Leaf quality of some tropical and temperate tree species as food resources for stream shredders. *International Reviews of Hydrobiology* **95**: 27–41.

Hallam A, Read J. 2006. Do tropical species invest more in anti-herbivore defence than temperate species? A test in *Eucryphia* (Cunoniaceae) in eastern Australia. *Journal of Tropical Ecology* 22: 41–51.

Hanley ME, Lamont BB, Fairbanks MM, Rafferty CM. 2007. Plant structural traits and their role in anti-herbivore defence. *Perspectives in Plant Ecology Evolution and Systematics* 8: 157–178.

Herms DA, Mattson WJ. 1992. The dilemma of plants: to grow or defend. *Quarterly Review of Biology* 67: 283–335.

Hodson MJ, White PJ, Mead A, Broadley MR. 2005. Phylogenetic variation in the silicon composition of plants. *Annals of Botany* 96: 1027–1046.

- Janzen DH. 1974. Tropical blackwater rivers, animals, and mast fruiting by the Dipterocarpaceae. *Biotropica* 6: 69–103.
- Jones AS, Lamont BB, Fairbanks MM, Rafferty CM. 2003. Kangaroos avoid eating seedlings with or near others with volatile essential oils. *Journal of Chemical Ecology* 29: 2621–2635.
- Jordan GJ, Dillon RA, Weston PH. 2005. Solar radiation as a factor in the evolution of scleromorphic leaf anatomy in Proteaceae. *American Journal of Botany* **92**: 789–796.

Kleidon A, Adams J, Pavlick R, Reu B. 2009. Simulated geographic variations of plant species richness, evenness and abundance using climatic constraints on plant functional diversity. *Environmental Research Letters* 4: 014007.

Korth KL, Doege SJ, Park SH, Goggin FL, Wang Q, Gomez SK, Liu G, Jia L, Nakata PA. 2006. *Medicago truncatula* mutants demonstrate the role of plant calcium oxalate crystals as an effective defense against chewing insects. *Plant Physiology* 141: 188–195.

Kursar TA, Coley PD. 1992. Delayed greening in tropical leaves – an antiherbivore defense. *Biotropica* 24: 256–262.

Lanning FC, Eleuterius LN. 1985. Silica and ash in tissues of some plants growing in the coastal area of Mississippi, USA. *Annals of Botany* 56: 157–172.

Lesage L, Crete M, Huot J, Ouellet JP. 2000. Quality of plant species utilized by northern white-tailed deer in summer along a climatic gradient. *Ecoscience* 7: 439–451.

Levin DA. 1976. Alkaloid-bearing plants: an ecogeographic perspective. *American Naturalist* 110: 261–284.

Levin DA, York BM. 1978. Toxicity of plant alkaloids: an ecogeographic perspective. *Biochemical Systematics and Ecology* 6: 61–76.

Lewinsohn TM. 1991. The geographical distribution of plant latex. Chemoecology 2: 64–68.

Lincoln DE. 1985. Host plant protein and phenolic resin effects on larval growth and survival of a butterfly. *Journal of Chemical Ecology* 11: 1459–1467.

MacArthur RH. 1972. Geographical ecology: patterns in the distribution of species. Princeton, NJ, USA: Princeton University Press.

Marko G, Gyuricza V, Bernath J, Altbacker V. 2008. Essential oil yield and composition reflect browsing damage of junipers. *Journal of Chemical Ecology* 34: 1545–1552. Marsh KJ, Wallis IR, Foley WJ. 2007. Behavioural contributions to the regulated intake of plant secondary metabolites in koalas. *Oecologia*, 154: 283–290.

Martz F, Peltola R, Fontanay S, Duval RE, Julkunen-Tiitto R, Stark S. 2009. Effect of latitude and altitude on the terpenoid and soluble phenolic composition of juniper (*Juniperus communis*) needles and evaluation of their antibacterial activity in the boreal zone. *Journal of Agricultural and Food Chemistry* 57: 9575–9584.

Massey FP, Ennos AR, Hartley SE. 2006. Silica in grasses as a defence against insect herbivores: contrasting effects on folivores and a phloem feeder. *Journal of Animal Ecology* 75: 595–603.

- Mattson WJ. 1980. Herbivory in relation to plant nitrogen content. Annual Review of Ecology and Systematics 11: 119–161.
- Moles AT, Bonser SP, Poore AGB, Wallis IR, Foley WJ. 2011. Assessing the evidence for latitudinal gradients in plant defence and herbivory. *Functional Ecology* 25: 380–388.
- Moles AT, Westoby M. 2000. Do small leaves expand faster than large leaves, and do shorter expansion times reduce herbivore damage? *Oikos* **90**: 517–524.
- Mueller-Harvey I. 2006. Unravelling the conundrum of tannins in animal nutrition and health. *Journal of the Science of Food and Agriculture* 86: 2010–2037.
- Ness JH, Morris WF, Bronstein JL. 2009. For ant-protected plants, the best defense is a hungry offense. *Ecology* **90**: 2823–2831.

O'Neill GA, Aitken SN, King JN, Alfaro RI. 2002. Geographic variation in resin canal defenses in seedlings from the Sitka spruce × white spruce introgression zone. *Canadian Journal of Forest Research-Revue Canadienne De Recherche Forestiere* 32: 390–400.

Peeters PJ. 2002. Correlations between leaf structural traits and the densities of herbivorous insect guilds. *Biological Journal of the Linnean Society* 77: 43–65.

- **Pemberton RW. 1998**. The occurrence and abundance of plants with extrafloral nectaries, the basis for antiherbivore defensive mutualisms, along a latitudinal gradient in east Asia. *Journal of Biogeography* 25: 661–668.
- Roslin T, Salminen JP. 2008. Specialization pays off: contrasting effects of two types of tannins on oak specialist and generalist moth species. *Oikos* 117: 1560–1568.
- Salminen J-P, Karonen M. 2011. Chemical ecology of tannins and other phenolics: we need a change in approach. *Functional Ecology* 25: 325–338.
- Schemske DW, Mittelbach GG, Cornell HV, Sobel JM, Roy K. 2009. Is there a latitudinal gradient in the importance of biotic interactions? *Annual Review of Ecology, Evolution and Systematics* 40: 245–269.
- Schmitz OJ. 2008. Herbivory from individuals to ecosystems. Annual Review of Ecology Evolution and Systematics 39: 133–152.

Shimada T. 2006. Salivary proteins as a defense against dietary tannins. Journal of Chemical Ecology 32: 1149–1163.

Silanikove N, Shinder D, Gilboa N, Eyal M, Nitsan Z. 1996. Binding of polyethylene glycol to samples of forage plants as an assay of tannins and their negative effects on ruminall degredation. *Journal of Agricultural* and Food Chemistry, 44: 3230–3234.

Siska EL, Pennings SC, Buck TL, Hanisak MD. 2002. Latitudinal variation in palatability of salt-marsh plants: which traits are responsible? *Ecology* 83: 3369–3381.

Spalinger DE, Collins WB, Hanley TA, Cassara NE, Carnahan AM. 2010. The impact of tannins on protein, dry matter, and energy digestion in moose (*Alces alces*). *Canadian Journal of Zoology-Revue Canadienne De Zoologie* 88: 977–987.

Stark S, Julkunen-Tiitto R, Holappa E, Mikkola K, Nikula A. 2008. Concentrations of foliar quercetin in natural populations of white birch (*Betula pubescens*) increase with latitude. *Journal of Chemical Ecology* 34: 1382–1391.

- Steinbauer MJ. 2010. Latitudinal trends in foliar oils of eucalypts: environmental correlates and diversity of chrysomelid leaf-beetles. *Austral Ecology* 35: 205–214.
- Turner IM. 1994. Sclerophylly primarily protective? *Functional Ecology* 8: 669–675.
- Van Alstyne KL, Dethier MN, Duggins DO. 2001. Spatial patterns in macroalgal chemical defenses. In: McClintock JB, Baker BJ, eds. *Marine chemical ecology*. New York, NY, USA: CRC Press, 301–324.

VanDam NM, DeJong TJ, Iwasa Y, Kubo T. 1996. Optimal distribution of defences: are plants smart investors? *Functional Ecology* 10: 128–136.

Venables WN, Ripley BD. 2002. *Modern applied statistics with S.* New York, NY, USA: Springer.

- Wainhouse D, Ashburner R, Ward E, Rose J. 1998. The effect of variation in light and nitrogen on growth and defence in young Sitka spruce. *Functional Ecology* 12: 561–572.
- Ward D, Spiegel M, Saltz D. 1997. Gazelle herbivory and interpopulation differences in calcium oxalate content of leaves of a desert lily. *Journal of Chemical Ecology* 23: 333–346.
- Webb LJ. 1959. A physiognomic classification of Australian rain forests. Journal of Ecology 47: 551–570.
- Wright IJ, Cannon K. 2001. Relationships between leaf lifespan and structural defences in a low-nutrient, sclerophyll flora. *Functional Ecology*, 15: 351–359.

Yamamura N, Tsuji N. 1995. Optimal strategy of plant antiherbivore defense – implications for apparency and resource-availability theories. *Ecological Research* 10: 19–30.

Supporting Information

Additional supporting information may be found in the online version of this article.

Table S1 Information about study sites

Table S2 Settings used to develop partial least squaresregression models to relate reflectance spectra from leafsamples to chemical traits

Table S3 Relationships between binary traits and latitude

Table S4 Relationships between species abundance andeach of the traits included in this study

Table S5 Results of phylogenetic analyses

Table S6 Analyses including a term for continent

Methods S1 Selection of sites, study species and individuals, near-infrared spectroscopy methods and phylogenetic analyses.

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